

## A BIPHENYL PHYTOALEXIN FROM *CERCIDIPHYLLUM JAPONICUM* \*

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**Key Word Index**—*Cercidiphyllum japonicum*; Cercidiphyllaceae; katsura tree; phytoalexin; magnolol; 5,5'-diallyl-2,2'-dihydroxybiphenyl.

**Abstract**—A biphenyl derivative has been isolated as a phytoalexin from the fungus-inoculated twig cortical tissue of *Cercidiphyllum japonicum*. The biphenyl was shown to be magnolol on the basis of spectral evidence and direct comparison with an authentic sample.

### INTRODUCTION

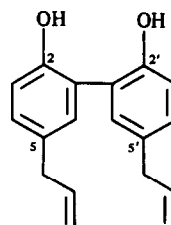
We have previously reported the isolation of several phytoalexins from moraceous trees [1, 2]. Screening tests for antifungal compounds from the fungus-inoculated tissues of other arboreal plants revealed that the twig cortical tissue of the katsura tree (*Cercidiphyllum japonicum*) produced a single antifungal compound (1), which was absent from water-treated control tissue. We report herein the isolation and characterization of this compound.

### RESULTS AND DISCUSSION

Bioassay-directed fractionation of the acetone extracts from the twig cortical tissue, which had been inoculated with *Fusarium solani* f. sp. *mori*, resulted in the isolation of the antifungal compound (1), which inhibited completely the conidial germination of *Bipolaris leersiae* at a concentration of  $5 \times 10^{-5}$  M. The chemical and spectral data of 1 suggested it to be 5,5'-diallyl-2,2'-dihydroxybiphenyl, magnolol. Direct comparison (mmp, IR and  $^1\text{H}$  NMR spectra) of 1 with those of magnolol confirmed its identity. This is the first report of magnolol as a phytoalexin although it has been reported as a constituent of *Magnolia* species [3, 4] and Taiwan sassafras [5]. To our knowledge, magnolol is the second biphenyl phytoalexin. The first one, aucuparin (4-hydroxy-3,5-dimethoxybiphenyl), was reported recently from two rosaceous plants *Eriobotrya japonica* [6] and *Malus pumila* [7].

### EXPERIMENTAL

**TLC bioassay.** Developed silica gel sheet (Merck, Kieselgel 60  $F_{254}$ ; ether) was air-dried, sprayed with a dense conidial suspension of *Bipolaris leersiae* in a potato-glucose medium, and incubated in a moist box at 25° for 2 days [8]. A fungitoxic area appeared white against a dark gray background.



1

**Induction and isolation of magnolol (1).** Twigs of *Cercidiphyllum japonicum* collected from Tomakomai Experiment Forest of Hokkaido University in September were stripped of epidermis with a razor-blade. The exposed cortical tissue was inoculated with a conidial suspension of *Fusarium solani* f. sp. *mori* and incubated in a moist chamber at 25° for 6 days. The browned tissue separated from the xylem was freeze-dried (523 g) and extracted with  $\text{Me}_2\text{CO}$  to afford the extracts (26.6 g) which showed a single antifungal spot ( $R_f$  0.51) on TLC bioassay. The extracts were chromatographed on a silica gel column using  $\text{CH}_2\text{Cl}_2$  with increasing amounts of MeOH. The combined active fractions (1.1 g) eluted with  $\text{CH}_2\text{Cl}_2$ -MeOH (100:20) were further separated on a Sephadex LH-20 column ( $\text{Me}_2\text{CO}$ ) and then on a silica gel column ( $\text{CH}_2\text{Cl}_2$ ) to afford a crystalline residue, which on recrystallization from  $\text{C}_6\text{H}_6$ -hexane gave 1 (226 mg); mp 100–101° (lit. 102°) [4],  $\text{C}_{18}\text{H}_{18}\text{O}_2$  (found:  $m/z$  266.1305);  $^1\text{H}$  NMR [500 MHz,  $(\text{CD}_3)_2\text{CO}$ ]:  $\delta$  3.36 (4H, br d,  $J = 6.7$  Hz,  $2 \times \text{CH}_2\text{CH}=\text{CH}_2$ ), 5.01 [2H, dddd,  $J = 10.1, 2.1, 1.2, 1.2$  Hz,  $2 \times \text{CH}_2\text{CH}=\text{CH}_2$  (cis)], 5.09 [2H, dddd,  $J = 17.1, 2.1, 1.5, 1.5$  Hz,  $2 \times \text{CH}_2\text{CH}=\text{CH}_2$  (trans)], 5.99 (2H, dddd,  $J = 17.1, 10.1, 6.7, 6.7$  Hz,  $2 \times \text{CH}_2\text{CH}=\text{CH}_2$ ), 6.92 (2H, d,  $J = 8.2$  Hz, H-3, H-3'), 7.07 (2H, dd,  $J = 8.2, 2.1$  Hz, H-4, H-4'), 7.11 (2H, d,  $J = 2.1$  Hz, H-6, H-6'), 8.24 (2H, OH);  $^{13}\text{C}$  NMR [25.15 MHz,  $(\text{CD}_3)_2\text{CO}$ ]: s at  $\delta$  127.0, 132.7, 153.0, d at  $\delta$  117.4, 129.6, 132.4, 139.0, t at  $\delta$  40.0, 115.5.

\* Part 4 in the series "Studies on Stress Metabolites". For part 3 see Takasugi, M., Kawashima, S., Monde, K., Katsui, N., Masamune, T. and Shirata, A., *Phytochemistry* (in press). Paper No. 6284

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